“BEST” PRACTICES FOR DISINFECTION OF NON-CRITICAL SURFACES AND EQUIPMENT AND MEDICAL WASTE MANAGEMENT

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LECTURE OBJECTIVES

- Review the CDC Guideline for Disinfection and Sterilization: Focus on role of environmental surfaces
- Review “best” practices for environmental cleaning and disinfection
- Review the use of low-level disinfectants and the activity of disinfectants on key hospital pathogens
- Review medical waste management
“BEST” PRACTICES

- There is limited scientific evidence to inform us on cleaning/disinfecting practices and frequency.
- There is little scientific evidence that disinfecting schedules should emphasize certain “high-risk” or “high-touch” sites.
ENVIRONMENTAL CONTAMINATION LEADS TO HAIs

- There is increasing evidence to support the contribution of the environment to disease transmission.

- This supports comprehensive disinfecting regimens (goal is not sterilization) to reduce the risk of acquiring a pathogen from the healthcare environment/equipment.
EH Spaulding believed that how an object will be disinfected depended on the object’s intended use:

- **CRITICAL** - objects which enter normally sterile tissue or the vascular system or through which blood flows should be sterile.

- **SEMICRITICAL** - objects that touch mucous membranes or skin that is not intact require a disinfection process (high-level disinfection [HLD]) that kills all microorganisms; however, small numbers of bacterial spores are permissible.

- **NONCRITICAL** - objects that touch only intact skin require low-level disinfection.
LECTURE OBJECTIVES

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- Review “best” practices for environmental cleaning and disinfection
- Review the use of low-level disinfectants and the activity of disinfectants on key hospital pathogens
- Review medical waste management
GUIDELINE FOR DISINFECTION
AND STERILIZATION IN
HEALTHCARE FACILITIES, 2008

Rutala WA, Weber DJ., HICPAC
Available on CDC web page-www.cdc.gov

William A. Rutala, Ph.D., M.P.H.\(^1,2\), David J. Weber, M.D., M.P.H.\(^1,2\), and the Healthcare Infection Control Practices Advisory Committee (HICPAC)\(^3\)
Blood Pressure Cuff
Non-Critical Patient Care Item
DISINFECTION OF NONCRITICAL PATIENT-CARE DEVICES

- Process noncritical patient-care devices using a disinfectant and concentration of germicide as recommended in the Guideline (IB)
- Disinfect noncritical medical devices (e.g., blood pressure cuff) with an EPA-registered hospital disinfectant using the label’s safety precautions and use directions. Most EPA-registered hospital disinfectants have a label contact time of 10 minutes but multiple scientific studies have demonstrated the efficacy of hospital disinfectants against pathogens with a contact time of at least 1 minute (IB)
- Ensure that, at a minimum noncritical patient-care devices are disinfected when visibly soiled and on a regular basis (e.g., once daily or weekly) (II)
- If dedicated, disposable devices are not available, disinfect noncritical patient-care equipment after using on a patient, who is on contact precautions before using this equipment on another patient (IB)
CLEANING AND DISINFECTION OF ENVIRONMENTAL SURFACES IN HEALTHCARE FACILITIES

- Clean housekeeping surfaces (e.g., floors, tabletops) on a regular basis, when spills occur, and when these surfaces are visibly soiled (II)
- Disinfect (or clean) environmental surfaces on a regular basis (e.g., daily, 3x per week) and when surfaces are visibly soiled (II)
- Follow manufacturers’ instructions for proper use of disinfecting (or detergent) products – such as recommended use-dilution, material compatibility, storage, shelf-life, and safe use and disposal (II)
- Clean walls, blinds, and window curtains in patient-care areas when these surfaces are visibly contaminated or soiled (II)
- Prepare disinfecting (or detergent) solutions as needed and replace with fresh solution frequently (e.g., replace floor mopping solution every 3 patient rooms, change no less often than at 60-minute intervals) (IB)
REVIEW THE "BEST" PRACTICES FOR CLEANING AND DISINFECTING

Cleaning and disinfecting is one-step with disinfectant-detergent. No pre-cleaning necessary unless spill or gross contamination. In many cases “best” practices not scientifically determined.
ALL “TOUCHABLE” (HAND CONTACT) SURFACES SHOULD BE WIPED WITH DISINFECTANT

“High touch” objects only recently defined (no significant differences in microbial contamination of different surfaces) and “high risk” objects not epidemiologically defined.
DAILY CLEANING/DISINFECTING PRACTICES

- Wash hands thoroughly and put on gloves
- Place wet floor sign at door
- Discard disposable items and remove waste and soiled linen
- Disinfect (damp wipe) all horizontal, vertical and contact surfaces with a cotton cloth saturated (or microfiber) with a disinfectant-detergent solution.
These surfaces (wipe all surfaces) include, but are not limited to:

- Bed rails
- Overbed table
- Infusion pumps
- IV poles/Hanging IV poles
- Nurse call box
- Monitor cables
- Telephone
- Countertops
DAILY CLEANING/DISINFECTING PRACTICES

● These surfaces include, but not limited to:
  ■ Soap dispenser
  ■ Paper towel dispenser
  ■ Cabinet fronts including handles
  ■ Visitor chair
  ■ Door handles inside and outside
  ■ Sharps container
  ■ TV remote, bed call remote
  ■ Bathroom-toilet seat, shower fixtures, flush handle
DAILY CLEANING/DISINFECTING PRACTICES

- Spot clean walls (when visually soiled) with disinfectant-detergent and windows with glass cleaner
- Clean and disinfect sink and toilet
- Stock soap and paper towel dispensers
- Damp mop floor with disinfectant-detergent
- Inspect work
- Remove gloves and wash hands
DAILY CLEANING/DISINFECTING PRACTICES

- Use EPA-registered disinfectant-detergent (if prepared on-site, document correct concentration)
- Cleaned surface should appear visibly wet and should be allowed to air dry at least one minute
- Change cotton mop water containing disinfectant every 3 rooms and after every isolation room
- Change cotton mop head after isolation room and after BBP spills (change microfiber after each room)
DAILY CLEANING/DISINFECTING PRACTICES

- Cleaning should be from the cleanest to dirtiest areas
  (the bathroom will be cleaned last followed by the floor)
- Change cleaning cloths after every room and use at least
  3 cloths per room; typically 5-7 cloths
- Do not place cleaning cloth back into the disinfectant
  solution after using it to wipe a surface
- Daily cleaning of certain patient equipment is the
  responsibility of other HCP (RC, nursing). Surfaces
  should be wiped with a clean cloth soaked in disinfectant
TERMINAL CLEANING/DISINFECTING PRACTICES

- “Terminal” or discharge cleaning of non-isolation rooms consists of the same procedure above plus disinfection of bed mattresses and inaccessible items
- Trash can cleaned weekly and when visible soiled
- Do not wash walls, strip and wax floors, or discard wrapped disposable supplies left in drawers
ISOLATION ROOM CLEANING

- ES staff use PPE required by the isolation card
- Same cleaning procedures as for non-isolation rooms (except C. difficile, norovirus)
- Do not use a dust mop or counter brush
- Leave the room only when completed (unless requested to leave by nurse or doctor)
A multidisciplinary team should develop C/D procedures for managing contamination.

A clean environment should be reestablished after the patient.
- Items used during surgical procedure should be C/D (OR beds, anesthesia carts/machines, positioning devices, Mayo stands).
- Floors/walls C/D after each surgical procedure if contaminated.

Terminal cleaning performed daily when used.
- C/D all exposed surfaces (anesthesia machine, OR beds, overhead lights, stools, positioning devices, etc).
SHOULD WE CONCENTRATE ON “HIGH TOUCH” OR “HIGH RISK” OBJECTS

No, not only “high risk” (all surfaces). “High touch” objects only recently defined and “high risk” objects not scientifically defined.
### Table: Rates of Cleaning for 14 Types of High-Risk Objects

<table>
<thead>
<tr>
<th>Object</th>
<th>Percentage cleaned</th>
<th>95% CI</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean ± SD</td>
<td>Range</td>
</tr>
<tr>
<td>Sink</td>
<td>82 ± 12</td>
<td>57-97</td>
</tr>
<tr>
<td>Toilet seat</td>
<td>76 ± 18</td>
<td>40-98</td>
</tr>
<tr>
<td>Tray table</td>
<td>77 ± 15</td>
<td>53-100</td>
</tr>
<tr>
<td>Bedside table</td>
<td>64 ± 22</td>
<td>23-100</td>
</tr>
<tr>
<td>Toilet handle</td>
<td>60 ± 22</td>
<td>23-89</td>
</tr>
<tr>
<td>Side rail</td>
<td>60 ± 21</td>
<td>25-96</td>
</tr>
<tr>
<td>Call box</td>
<td>50 ± 19</td>
<td>9-90</td>
</tr>
<tr>
<td>Telephone</td>
<td>49 ± 16</td>
<td>18-86</td>
</tr>
<tr>
<td>Chair</td>
<td>48 ± 28</td>
<td>11-100</td>
</tr>
<tr>
<td>Toilet door knobs</td>
<td>28 ± 22</td>
<td>0-82</td>
</tr>
<tr>
<td>Toilet hand hold</td>
<td>28 ± 23</td>
<td>0-90</td>
</tr>
<tr>
<td>Bedpan cleaner</td>
<td>25 ± 18</td>
<td>0-79</td>
</tr>
<tr>
<td>Room door knobs</td>
<td>23 ± 19</td>
<td>2-73</td>
</tr>
<tr>
<td>Bathroom light switch</td>
<td>20 ± 21</td>
<td>0-81</td>
</tr>
</tbody>
</table>

**Note:** CI, confidence interval.
“The patient in the next bed is highly infectious. Thank God for these curtains.”
Hospital Privacy Curtains
(sprayed “grab area” 3x from 6-8” with 1.4% IHP and allowed 2 minute contact; sampled)
Decontamination of Curtains with Activated HP (1.4%)

<table>
<thead>
<tr>
<th>CP for:</th>
<th>Before Disinfection CFU/5 Rodacs (#Path)</th>
<th>After Disinfection CFU/5 Rodacs (#Path)</th>
<th>% Reduction</th>
</tr>
</thead>
<tbody>
<tr>
<td>MRSA</td>
<td>330 (10 MRSA)</td>
<td>21*(0 MRSA)</td>
<td>93.6%</td>
</tr>
<tr>
<td>MRSA</td>
<td>186 (24 VRE)</td>
<td>4* (0 VRE)</td>
<td>97.9%</td>
</tr>
<tr>
<td>MRSA</td>
<td>108 (10 VRE)</td>
<td>2* (0 VRE)</td>
<td>98.2%</td>
</tr>
<tr>
<td>VRE</td>
<td>75 (4 VRE)</td>
<td>0 (0 VRE)</td>
<td>100%</td>
</tr>
<tr>
<td>VRE</td>
<td>68 (2 MRSA)</td>
<td>2* (0 MRSA)</td>
<td>97.1%</td>
</tr>
<tr>
<td>VRE</td>
<td>98 (40 VRE)</td>
<td>1* (0 VRE)</td>
<td>99.0%</td>
</tr>
<tr>
<td>MRSA</td>
<td>618 (341 MRSA)</td>
<td>1* (0 MRSA)</td>
<td>99.8%</td>
</tr>
<tr>
<td>MRSA</td>
<td>55 (1 VRE)</td>
<td>0 (0 MRSA)</td>
<td>100%</td>
</tr>
<tr>
<td>MRSA, VRE</td>
<td>320 (0 MRSA, 0 VRE)</td>
<td>1* (0 MRSA, 0 VRE)</td>
<td>99.7%</td>
</tr>
<tr>
<td>MRSA</td>
<td>288 (0 MRSA)</td>
<td>1* (0 MRSA)</td>
<td>99.7%</td>
</tr>
<tr>
<td>Mean</td>
<td>2146/10=215 (432/10=44)</td>
<td>33*/10=3 (0)</td>
<td>98.5%</td>
</tr>
</tbody>
</table>

* All isolates after disinfection were *Bacillus sp*; now treat CP patient curtains at discharge
TERMINAL CLEANING PRACTICE

- Some hospitals change curtains after Contact Precaution patients.
- At UNC Health Care, privacy curtains are changed routinely every 3 months or when visibly soiled.
- In Contact Precaution rooms, frequently touched surfaces of the curtains should be sprayed with approved disinfectant (e.g., improved HP).
- Vinyl shower curtains are cleaned when visibly soiled or replaced as needed.
Cleaning/Disinfection

- ES and nursing need to agree on who is responsible for cleaning what (especially equipment)

- ES needs to know
  - Which disinfectant/detergent to use
  - What concentration would be used (and verified)
  - What contact times are recommended (bactericidal)
  - How often to change cleaning cloths/mop heads
  - How important their job is to infection prevention
LECTURE OBJECTIVES

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DISINFECTING NONCRITICAL PATIENT EQUIPMENT AND ENVIRONMENTAL SURFACES

Classification: Noncritical objects will not come in contact with mucous membranes or skin that is not intact.

Object: Can be expected to be contaminated with some microorganisms.

Level germicidal action: Kill vegetative bacteria, fungi and lipid viruses.

Examples: Bedpans; crutches; bed rails; EKG leads; bedside tables; walls, floors and furniture.

Method: Low-level disinfection
Low-Level Disinfection for Noncritical Equipment and Surfaces

Exposure time ≥ 1 min

<table>
<thead>
<tr>
<th>Germicide</th>
<th>Use Concentration</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ethyl or isopropyl alcohol</td>
<td>70-90%</td>
</tr>
<tr>
<td>Chlorine</td>
<td>100ppm (1:500 dilution)</td>
</tr>
<tr>
<td>Phenolic</td>
<td>UD</td>
</tr>
<tr>
<td>Iodophor</td>
<td>UD</td>
</tr>
<tr>
<td>Quaternary ammonium (QUAT)</td>
<td>UD</td>
</tr>
<tr>
<td>QUAT with alcohol</td>
<td>RTU</td>
</tr>
<tr>
<td>Improved hydrogen peroxide (HP)</td>
<td>0.5%, 1.4%</td>
</tr>
<tr>
<td>Peracetic acid with HP (C. difficile)</td>
<td>UD</td>
</tr>
</tbody>
</table>

UD=Manufacturer’s recommended use dilution; others in development/testing-electrolyzed water; polymeric guanidine; cold-air atmospheric pressure plasma (Boyce Antimicrob Res IC 2016. 5:10)
Decreasing Order of Resistance of Microorganisms to Disinfectants/Sterilants

Most Resistant
- Prions
- Spores (*C. difficile*)
- Mycobacteria
- Non-Enveloped Viruses (*norovirus*)
- Fungi
- Bacteria (*MRSA, VRE, Acinetobacter*)

Most Susceptible
- Enveloped Viruses
Most prevalent pathogens causing HAI (~75% easy to kill)

- S. aureus (15.6%)
- E. coli (11.5%)
- Coag neg Staph (11.4%)
- Klebsiella (8.0%)
- P. aeruginosa (8.0%)
- E. faecalis (6.8%)
- C. albicans (5.3%)
- Enterobacter sp. (4.7%)
- Other Candida sp (4.2%)
- C. difficile in top 2-3 past 5 years

Common causes of outbreaks and ward closures (relatively hard to kill)

- C. difficile spores
- Norovirus
- Rotavirus
- Adenovirus
MOST PREVALENT PATHOGENS
CAUSING HAI

Most prevent pathogens causing HAI (easy to kill)
- S. aureus (15.6%)
- E. coli (11.5%)
- Coag neg Staph (11.4%)
- Klebsiella (8.0%)
- P. aeruginosa (8.0%)
- E. faecalis (6.8%)
- C. albicans (5.3%)
- Enterobacter sp. (4.7%)
- Other Candida sp (4.2%)

Common causes of outbreaks and ward closures (relatively hard to kill)
- C. difficile spores
- Norovirus
- Rotavirus
- Adenovirus
## LOW-LEVEL DISINFECTION FOR NONCRITICAL EQUIPMENT AND SURFACES

Exposure time $\geq$ 1 min

<table>
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<th>Use Concentration</th>
</tr>
</thead>
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<td>Ethyl or isopropyl alcohol</td>
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<td>Phenolic</td>
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</tr>
<tr>
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UD=Manufacturer’s recommended use dilution; others in development/testing-electrolyzed water; polymeric guanidine; cold-air atmospheric pressure plasma (Boyce Antimicrob Res IC 2016. 5:10)
PROPERTIES OF AN IDEAL DISINFECTANT

● Broad spectrum-wide antimicrobial spectrum
● Fast acting-should produce a rapid kill
● Remains Wet-meet listed kill/contact times with a single application
● Not affected by environmental factors-active in the presence of organic matter
● Nontoxic-not irritating to user
● Surface compatibility-should not corrode instruments and metallic surfaces
● Persistence-should have sustained antimicrobial activity
● Easy to use
● Acceptable odor
● Economical-cost should not be prohibitively high
● Soluble (in water) and stable (in concentrate and use dilution)
● Cleaner (good cleaning properties) and nonflammable
# Key Considerations for Selecting the Ideal Disinfectant for Your Facility


<table>
<thead>
<tr>
<th>Consideration</th>
<th>Question to Ask</th>
<th>Score (1-10)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Kill Claims</td>
<td>Does the product kill the most prevalent healthcare pathogens.</td>
<td></td>
</tr>
<tr>
<td>Kill Times and Wet-Contact Times</td>
<td>How quickly does the product kill the prevalent healthcare pathogens. Ideally, contact time greater than or equal to the kill claim.</td>
<td></td>
</tr>
<tr>
<td>Safety</td>
<td>Does the product have an acceptable toxicity rating, flammability rating.</td>
<td></td>
</tr>
<tr>
<td>Ease-of-Use</td>
<td>Odor acceptable, shelf-life, in convenient forms (wipes, spray), water soluble, works in organic matter, one-step (cleans/disinfects)</td>
<td></td>
</tr>
<tr>
<td>Other factors</td>
<td>Supplier offer comprehensive training/education, 24-7 customer support, overall cost acceptable (product capabilities, cost per compliant use, help standardize disinfectants in facility)</td>
<td></td>
</tr>
</tbody>
</table>

Note: Consider the 5 components shown, give each product a score (1 is worst and 10 is best) in each of the 5 categories, and select the product with the highest score as the optimal choice (maximum score is 50).
Quaternary ammonium compounds
(e.g., didecyl dimethyl ammonium bromide, dioctyl dimethyl ammonium bromide)

## Advantages
- Bactericidal, fungicidal, virucidal against enveloped viruses (e.g., HIV)
- Good cleaning agents
- EPA registered
- Surface compatible
- Persistent antimicrobial activity when undisturbed
- Inexpensive (in dilutable form)
- Not flammable

## Disadvantages
- Not sporicidal
- In general, not tuberculocidal and virucidal against non-enveloped viruses
- High water hardness and cotton/gauze can make less micbicidal
- A few reports documented asthma as result of exposure to benzalkonium chloride
- Affected by organic matter
- Multiple outbreaks ascribed to contaminated benzalkonium chloride
### Alcohol

**Advantages**
- Bactericidal, tuberculocidal, fungicidal, virucidal
- Fast acting
- Non-corrosive
- Non-staining
- Used to disinfect small surfaces such as rubber stoppers on medication vials
- No toxic residue

**Disadvantages**
- Not sporicidal
- Affected by organic matter
- Slow acting against non-enveloped viruses (e.g., norovirus)
- No detergent or cleaning properties
- Not EPA registered
- Damage some instruments (e.g., harden rubber, deteriorate glue)
- Flammable (large amounts require special storage)
- Evaporates rapidly making contact time compliance difficult
- Not recommended for use on large surfaces
- Outbreaks ascribed to contaminated alcohol
Adenovirus is a hardy virus that is relatively resistant to disinfectants

- Quat about $<0.5 \log_{10}$ reduction against adenovirus with 1m exposure time
- Accelerated hydrogen peroxide (0.5%) demonstrates $\sim0.7 \log_{10}$ reduction against adenovirus with 1m exposure time
- Quat/Alcohol demonstrates a $\sim4 \log_{10}$ reduction against adenovirus with 1m exposure time
- Chlorine (~5000ppm) demonstrates a $\sim5 \log_{10}$ reduction against adenovirus with 1m exposure time
- Quat/Alcohol has improved virucidal activity compared to Quat and accelerated hydrogen peroxide
Improved Hydrogen Peroxide

Advantages
- Bactericidal, tuberculocidal, fungicidal, virucidal
- Fast efficacy
- Easy compliance with wet-contact times
- Safe for workers (lowest EPA toxicity category, IV)
- Benign for the environment
- Surface compatible
- Non-staining
- EPA registered
- Not flammable

Disadvantages
- More expensive than most other disinfecting actives
- Not sporicidal at low concentrations
**Sodium Hypochlorite**


<table>
<thead>
<tr>
<th>Advantages</th>
<th>Disadvantages</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bactericidal, tuberculocidal, fungicidal, virucidal</td>
<td>Reaction hazard with acids and ammonias</td>
</tr>
<tr>
<td>Sporicidal</td>
<td>Leaves salt residue</td>
</tr>
<tr>
<td>Fast acting</td>
<td>Corrosive to metals (some ready-to-use products may be formulated with</td>
</tr>
<tr>
<td></td>
<td>corrosion inhibitors)</td>
</tr>
<tr>
<td>Inexpensive (in dilutable form)</td>
<td>Unstable active (some ready-to-use products may be formulated with</td>
</tr>
<tr>
<td></td>
<td>stabilizers to achieve longer shelf life)</td>
</tr>
<tr>
<td>Not flammable</td>
<td>Affected by organic matter</td>
</tr>
<tr>
<td>Unaffected by water hardness</td>
<td>Discolors/stains fabrics</td>
</tr>
<tr>
<td>Reduces biofilms on surfaces</td>
<td>Potential hazard is production of trihalomethane</td>
</tr>
<tr>
<td>Relatively stable (e.g., 50% reduction in chlorine concentration in 30 days)</td>
<td>Odor (some ready-to-use products may be formulated with odor inhibitors).</td>
</tr>
<tr>
<td></td>
<td>Irritating at high concentrations.</td>
</tr>
<tr>
<td>Used as the disinfectant in water treatment</td>
<td></td>
</tr>
<tr>
<td>EPA registered</td>
<td></td>
</tr>
</tbody>
</table>
Phenolics

Advantages
- Bactericidal, tuberculocidal, fungicidal, virucidal
- Inexpensive (in dilutable form)
- Non-staining
- Not flammable
- EPA registered

Disadvantages
- Not sporidical
- Absorbed by porous materials and irritate tissue
- Depigmentation of skin caused by certain phenolics
- Hyperbilirubinemia in infants when phenolic not prepared as recommended
ALL “TOUCHABLE” (HAND CONTACT) SURFACES SHOULD BE WIPED WITH DISINFECTANT

“High touch” objects only recently defined (no significant differences in microbial contamination of different surfaces) and “high risk” objects not epidemiologically defined.
Effective Surface Decontamination

Product and Practice = Perfection
MICROBIAL BURDEN ON ROOM SURFACES AS A FUNCTION OF FREQUENCY OF TOUCHING

<table>
<thead>
<tr>
<th>Surface</th>
<th>Prior to Cleaning</th>
<th>Post Cleaning (mean)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean CFU/RODAC (95% CI)</td>
<td>Mean CFU/RODAC (95% CI)</td>
</tr>
<tr>
<td>High</td>
<td>71.9 (46.5-97.3)</td>
<td>9.6</td>
</tr>
<tr>
<td>Medium</td>
<td>44.2 (28.1-60.2)</td>
<td>9.3</td>
</tr>
<tr>
<td>Low</td>
<td>56.7 (34.2-79.2)</td>
<td>5.7</td>
</tr>
</tbody>
</table>

- The level of microbial contamination of room surfaces is similar regardless of how often they are touched both before and after cleaning.
- Therefore, all surfaces that are touched must be cleaned and disinfected.
Thoroughness of Environmental Cleaning

Carling P. AJIC 2013;41:S20-S25

Mean = 32%

Mean = 32%

DAILY CLEANING
TERMINAL CLEANING

>110,000
Objects

| = 95% CI
Wipes

Cotton, Disposable, Microfiber, Cellulose-Based, Nonwoven Spunlace
WIPES

- Wipes-cotton, disposable, microfiber, nonwoven spunlace
- Wipe should have sufficient wetness to achieve the disinfectant contact time. Discontinue use of the wipe if no longer leaves the surface visible wet for \( \geq 1 \) minute.
- When the wipe is visibly soiled, flip to a clean/unused side and continue until all sides of the wipe have been used (or get another wipe)
- Dispose of the wipe/cloth wipe appropriately
- Do not re-dip a wipe into the clean container of pre-saturated wipes
DISPOSABLE WIPES


- Wetness-ideally, stays wet long enough to meet EPA-registered contact times (e.g., bacteria-1 minute).
- Surface Coverage-premoistened wipe keeps surface area wet for 1-2 minutes (e.g., 12”x12” wipes keep 55.5 sq ft wet for 2m; 6”x5” equipment wipe keeps 6.7 sq ft wet for 2m). Wipe size based on use from small surfaces to large surfaces like mattress covers
- Durable substrate-will not easily tear or fall apart
- Top-keep closed or wipes dry out
Surface Disinfection:
Treatment Time (Wipes/Sprays) versus Contact Time (Liquids)

Dilutable liquid disinfectant-contact time is “wet” time

Wipes/Sprays-treatment time is undisturbed time ("wet” time is not relevant)
Surface Disinfection:  
Treatment Time (Wipes/Sprays) versus Contact Time (Liquids)  
Rutala, Weber. Submitted for publication

- Registration test for liquid disinfectants is the AOAC Use-Dilution Method (UDM).
- SS cylinders are inoculated with the test organism (S. aureus, S. choleraesuis, P. aeruginosa) and then dried. After drying, the cylinder is transferred to a disinfectant tube and immersed in the disinfectant for the contact time (e.g., 5 minutes).
- Thus, for liquid disinfectants tested by the UDM, the contact time should be the “wet” time (not undisturbed time).
Surface Disinfection:

Treatment Time (Wipes/Sprays) versus Contact Time (Liquids)


- Registration test for wipe is EPA Disinfectant Towelette Test
- Treatment time is equal to combination of physical removal and inactivation caused by the disinfectant regardless of the surface appearance (i.e. wet or dry)
- Thus, if disinfectant wipe has a registration time of 1 minute, then the surface should be allowed to remain undisturbed for the registration time of 1 minute (i.e. wet time is not relevant)
LECTURE OBJECTIVES

- Review the CDC Guideline for Disinfection and Sterilization: Focus on role of environmental surfaces
- Review “best” practices for environmental cleaning and disinfection
- Review the use of low-level disinfectants and the activity of disinfectants on key hospital pathogens
- Review medical waste management
Medical Waste Problem

- Perceived threat of AIDS via medical waste
- Beach wash-ups of “medical waste”
- Overly restrictive medical waste rules and increase in volume of regulated medical waste
- Options for medical waste treatment and disposal diminishing
Definitions

Hospital waste (solid waste) refers to all waste (biological or nonbiological) which are discarded and not intended for further use (e.g., administrative waste, dietary waste)

Medical waste refers to materials generated as a result of patient diagnosis, treatment, or immunization (e.g., soiled dressing, intravenous tubing)

Regulated medical waste (“infectious” waste) refers to that portion of medical waste which could transmit an infectious disease (e.g., microbiological waste, sharps)
# Total Hospital Waste Generated per Patient by Bed Size

Rutala, Odette, Samsa. JAMA;262:1635-1640

<table>
<thead>
<tr>
<th>Hospital Beds</th>
<th>Ib/Bed/Day</th>
<th>Median</th>
<th>Ib/Patient/Day</th>
<th>Median</th>
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</thead>
<tbody>
<tr>
<td></td>
<td>N</td>
<td>Total</td>
<td>N</td>
<td>Total</td>
</tr>
<tr>
<td>&lt;100</td>
<td>90</td>
<td>5.72</td>
<td>69</td>
<td>11.30</td>
</tr>
<tr>
<td>100-299</td>
<td>108</td>
<td>10.36</td>
<td>84</td>
<td>15.79</td>
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<tr>
<td>300-499</td>
<td>40</td>
<td>12.51</td>
<td>32</td>
<td>18.47</td>
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<tr>
<td>&gt;500</td>
<td>27</td>
<td>12.86</td>
<td>23</td>
<td>16.95</td>
</tr>
<tr>
<td>Combined</td>
<td>265</td>
<td>9.21</td>
<td>208</td>
<td>15.28</td>
</tr>
</tbody>
</table>
Regulated Medical Waste

The CDC, EPA, and states define medical waste as regulated (“infectious”)

When it is suspected to contain potentially hazardous levels of microorganisms
Factors Necessary for Induction of Disease

- Dose
- Resistance of host
- Portal of entry
- Presence of a pathogen
- Virulence
Medical Waste

Plausible Transmission Routes


- Virtually nonexistent - respiratory, urinary or gastrointestinal tract or mucous membrane of the mouth, eyes, nose.
- Why? Waste must contain pathogens -> person must come in direct contact -> inject, ingest, or injury must follow the contact thereby creating portal of entry -> an infectious dose must enter susceptible host via portal of entry -> agent causes infection.
- Rare - "Sharps" have an intrinsic capability to disrupt the skin's integrity and introduce infectious agents.
Epidemiologic Evidence

• Only medical waste associated with infectious disease transmission is contaminated sharps.
• All reports of transmission of infectious agents by sharps occurred in health care setting.
• No evidence that a member of the public or a waste industry worker has ever acquired infection from medical waste (one exception).
• No infectious risks associated with any type of medical waste treatment method to include sanitary landfill disposal.
Microbiologic Quality

- Household waste contains on average 100x more microorganisms with pathogenic potential for humans than medical waste.

- Common nosocomial pathogens (i.e. *P. aeruginosa*, *Klebsiella* spp, *Enterobacter* spp, *Proteus* spp) were detected more frequently from household waste than from hospital waste.
Regulated Medical Waste

The CDC, EPA, and states define medical waste as regulated ("infectious") when it is suspected to contain potentially hazardous levels of microorganisms.
Medical Waste Regulations

- State - designation, transportation, storage and treatment
- Federal (OSHA) - education, labeling, use of PPE
# Types of Solid Waste Designated as Infectious and Recommended Disposal Methods


## Centers for Disease Control

<table>
<thead>
<tr>
<th>Source/Type</th>
<th>Infectious Waste</th>
<th>Disposal Method</th>
</tr>
</thead>
<tbody>
<tr>
<td>Microbiological</td>
<td>Yes</td>
<td>S, I</td>
</tr>
<tr>
<td>Blood and blood products</td>
<td>Yes</td>
<td>S, I, Sew</td>
</tr>
<tr>
<td>Pathological</td>
<td>Yes</td>
<td>Yes</td>
</tr>
<tr>
<td>Sharps (especially needles)</td>
<td>Yes</td>
<td>S, I</td>
</tr>
<tr>
<td>Contaminated animal carcasses (carcasses)</td>
<td>Yes</td>
<td>S, I</td>
</tr>
<tr>
<td>Isolation</td>
<td>No</td>
<td>—</td>
</tr>
<tr>
<td>Other (surgical waste, dialysis, contaminated lab waste)</td>
<td>No</td>
<td>—</td>
</tr>
</tbody>
</table>

Abbreviations: S-steam; I-incineration; Sew-sanitary sewer.
# Compliance with CDC and EPA Recommendations for Treatment of Regulated Medical Waste

Rutala, Odette, Samsa. JAMA;262:1635-1640

<table>
<thead>
<tr>
<th>Type of Medical Waste</th>
<th>CDC</th>
<th>USH%</th>
<th>EPA</th>
<th>USH%</th>
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<tbody>
<tr>
<td>Microbiological</td>
<td>S,I</td>
<td>98.1</td>
<td>S,I,T1,C</td>
<td>98.1</td>
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<tr>
<td>Blood</td>
<td>S,I,SEW</td>
<td>95.9</td>
<td>S,I,SEW,C</td>
<td>95.9</td>
</tr>
<tr>
<td>Pathology</td>
<td>I</td>
<td>92.6</td>
<td>I,SW,CB</td>
<td>92.6</td>
</tr>
<tr>
<td>Sharps</td>
<td>S,I</td>
<td>92.5</td>
<td>S,I</td>
<td>92.5</td>
</tr>
<tr>
<td>Isolation</td>
<td>—</td>
<td>—</td>
<td>S,I</td>
<td>85.9</td>
</tr>
<tr>
<td>Cont. animal carcasses</td>
<td>I</td>
<td>89.1</td>
<td>I,SW</td>
<td>89.1</td>
</tr>
<tr>
<td>Contaminated laboratory</td>
<td>—</td>
<td>—</td>
<td>Optional</td>
<td>87.0</td>
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<tr>
<td>Surgery</td>
<td>—</td>
<td>—</td>
<td>Optional</td>
<td>78.2</td>
</tr>
<tr>
<td>Autopsy</td>
<td>—</td>
<td>—</td>
<td>Optional</td>
<td>89.9</td>
</tr>
<tr>
<td>Dialysis</td>
<td>—</td>
<td>—</td>
<td>Optional</td>
<td>68.6</td>
</tr>
<tr>
<td>Contaminated equipment</td>
<td>—</td>
<td>—</td>
<td>Optional</td>
<td>ND</td>
</tr>
<tr>
<td>Overall</td>
<td>—</td>
<td>—</td>
<td></td>
<td>82.3</td>
</tr>
<tr>
<td>Overall</td>
<td>—</td>
<td>—</td>
<td></td>
<td>75.1</td>
</tr>
</tbody>
</table>
Regulated Waste:

OSHA

- Contaminated sharps
- Pathological and microbiological wastes containing blood or OPIM
- Liquid or semi-liquid blood or OPIM
- Contaminated items that would release blood or OPIM in a liquid or semi-liquid state if compressed
- Items caked with dried blood or OPIM that are capable of releasing these materials during handling
Regulated Waste:
OSHA

Can OSHA and states adopt uniform definitions of RMW?

- OSHA rules and state rules address two different concerns
  - OSHA rule addresses waste management in the workplace to ensure worker safety
  - State waste management rules ensure storage, shipping, and treatment/disposal practices that protect the environment and public health
Segregation of Medical Waste by US Hospitals

95% segregate regulated medical waste from non-regulated medical waste

96% use labeled or color-coded bags
Collection and Containment of US Hospital Waste

Collection

Housekeeping (82%), maintenance (4%)
or both (7%) transfer wastes to on-site
storage or processing site (at least daily) 92%

Container

Leakproof wastebaskets 95%
Plastic bags as wastebasket liners 99%
Transporting Waste Within US Hospitals

Transfer Carts - used to transport waste within the hospital 95%
Gravity Chutes - allows for vertical transfer 13%
Pneumatic chutes - vacuum source to propel wastes 2%
Storage

- No single requirement for storage of infectious waste in terms of time and temperature but most states do have regulations.

- NC - if not shipped within seven days of generation, medical waste must be refrigerated.
Compaction

● Advantages
  ■ Decreases the volume of waste 4-5 times
  ■ Decreases size of storage facilities
  ■ Decreases cost of transporting waste

● Disadvantage
  ■ May interfere with the effectiveness of certain treatment processes
Medical Waste Management:
Environmentally Responsible Healthcare

• Treatment of regulated medical waste (RMW)
  • Properly define RMW
  • Rational definition could save millions in treatment costs
  • Reduce waste now treated as RMW (e.g., incineration)
    • Know what happens to facility’s wastes and how treated
    • Locate “red bags” strategically to capture RMW
  • Encourage segregation of properly defined RMW
### Treatment of Medical Waste by US Hospitals

Rutala, Odette, Samsa. JAMA;262:1635-1640

<table>
<thead>
<tr>
<th>Waste Category</th>
<th>Infectious (%)</th>
<th>Treatment/Disposal Methods (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Yes</td>
<td>No</td>
</tr>
<tr>
<td>Microbiological</td>
<td>99</td>
<td>1</td>
</tr>
<tr>
<td>Human blood</td>
<td>94</td>
<td>6</td>
</tr>
<tr>
<td>Pathological</td>
<td>96</td>
<td>4</td>
</tr>
<tr>
<td>Isolation</td>
<td>94</td>
<td>6</td>
</tr>
<tr>
<td>Sharps</td>
<td>99</td>
<td>1</td>
</tr>
</tbody>
</table>
Incineration

Ash Residue: Controlled-air incinerators produce a sterile ash

Air Emissions: No difference between bacteria in stack emissions and ambient air. Chemicals (CO, metals, acid gases, dioxins, furans) emitted from hospital or municipal incinerators
EPA’s Proposed Incineration Emission Limits

- New set of medical waste incineration regulations
- Regulates Hosp/Med/Inf Waste Incinerators
- Mandated by the Clean Air Act Amendments of 1990
- Regulations will substantially reduce emissions (dioxins, CO, Pb, Hg)
- EPA estimates regulations would close 50-80% of existing medical waste incinerators.
Steam Sterilization

- 250°F for 45 min or other effective combination.
- Unit should have time-temperature recorder and pressure gauge.
- Biological monitoring at least weekly; log maintained and shall include type of indicator used, date, time and result of test.
Sanitary Landfill

- Untreated medical waste could be discarded in sanitary landfills provided workers do not have contact.

- Studies demonstrate:
  - Bacteria and viruses are reduced by thermal inactivation, antimicrobial characteristics of leachate and absorption to organic material.
  - Household waste is more microbially contaminated and it is discarded in sanitary landfills.

- Unavailable, reaching capacity, or restricted to untreated medical waste.
Needle Disposal

“Needles should not be recapped, purposely bent or broken by hand, removed from disposal syringes, or otherwise manipulated by hand. After they are used, disposable syringes and needles, scalpel blades, and other sharp items should be placed in puncture-resistant containers for disposal; the puncture-resistant containers should be located as close as practical to the use area.”

Centers for Disease Control, MMWR August 21, 1987
Suctioned Fluids Disposal Treatment

Sanitary sewer - suctioned fluids may be carefully poured down a drain connected to a sanitary sewer (CDC, 1985)

Incineration - suction canister may be tightly capped, bagged (coded) and incinerated.

Sanitary landfill - suction canister fluid may be treated with liquid treatment system and sent to sanitary landfill (2/3 states)

Sanitary landfill - suction canisters may be tightly capped, bagged and sent to a sanitary landfill (a few states).
Medical Waste Management: Treatment

● Goal of treatment is to reduce microbial load

● Changes that impact treatment of RMW
  ■ Incineration (new emission standards)
  ■ Alternative treatment technologies
    - Some states allow alternatives, other states not
  ■ Autoclave
Medical Waste Management:
Treatment

● Alternative technologies
  ■ Volume of medical waste reduced by equipment (grinders, shredders, hammermills) that pulverize and shred waste
  ■ Reduce volume by about 80%
  ■ Disinfect in the process (e.g. chemicals, heat, steam, microwaves, electrothermal radiation)
Alternative Medical Waste Treatment Technologies

- Microwaving - thermal decontamination
- Electro-thermal radiation - macrowaves
- Infrared heating
- Pyrolysis - heating (1000°F-6000°F) in absence of oxygen
- Plasma burners - electrical arcs used to create plasmas in range of 2500°C to 10000°C
Alternative Medical Waste Treatment Technologies (cont)

- Chemical processes
  - Bleach decontamination with shredding
  - Shredding followed by chlorine dioxide treatment
  - Shredding followed by wet oxidation at 212°F, pH 0.
  - Polymers with disinfectants solidify and decontaminate

- Irradiation processes
  - Gamma radiation such as cobalt 60
  - Electron beam radiation

Adapted from ASHMM handout by Lawrence G. Doucet, P.E.
Medical Waste Regulations

- State - designation, transportation, storage and treatment

- Federal (OSHA) - education, labeling, use of PPE
North Carolina Medical Waste Rules

Regulated Medical Waste Definitions

Microbiological - cultures and stocks of infectious agents

Pathological - human tissues, organs and body parts; carcasses and body parts of animals exposed to pathogens

Blood - liquid blood, serum, plasma, other blood products, emulsified human tissue, spinal fluids, and pleural and peritoneal fluids; in individual containers in volumes greater than 20 ml (bloody gauze, used gloves, tubing and dressings are not regulated medical waste).
**North Carolina Medical Waste Rules**

- **Definition** - “sharps” means and includes needles, syringes with attached needles, capillary tubes, slides, cover slips and scalpel blades.

- **Requirement** - sharps will be placed in a container which is rigid, leakproof when in an upright position and puncture-resistant. Contained sharps shall not be compacted prior to off-site transportation.

- **Treatment** - none required. The package may be disposed with general solid waste.
North Carolina Medical Waste Rules

Regulated Medical Waste Treatment*

Microbiological - incineration, steam sterilization or chemical treatment

Pathological - incineration

Blood and body fluids in individual containers in volumes greater than 20 ml - incineration or sanitary sewage systems, provided the sewage treatment authority is notified.

*Other methods of treatment shall require approval by the Division of Solid Waste Management
Medical Waste Minimization

Recycling/Reuse/Reduction

- Corrugated boxes and paper products
- Aluminum, glass, and plastic from defined areas
- Recovery/redistillation of laboratory solvents (e.g., alcohol, xylene, toluene)
- Source reduction-replace single use items with reusable items
Infectious Risks Associated with Recycling Hospital Waste

- No infectious risks associated with recycling hospital waste
- Presently, recycling efforts have generally focused on nonpatient contact sources of waste such as glass, scrap metal, aluminum cans, cardboard and packaging material
- From an infectious disease perspective, only a few items generated in the health-care setting are not likely candidates for recycling (e.g. sharps)
Waste Management:
How to be Friendly to the Environment

Recycling in healthcare

- Internal forces: employee requests, environment, public image, proactive posture
- External forces: state/national solid waste laws, local government regulations, air quality regulations
- Example: NC
  - 1989-GS established recycling goal of 25% by 1993
  - 1991-Amended to waste reduction and 40% by 2001
  - 1995-Amended so County government selects own goal
  - Orange county selected a reduction goal of 45%
LECTURE OBJECTIVES

- Review the CDC Guideline for Disinfection and Sterilization: Focus on role of environmental surfaces
- Review “best” practices for environmental cleaning and disinfection
- Review the use of low-level disinfectants and the activity of disinfectants on key hospital pathogens
- Review medical waste management
BEST PRACTICES FOR SURFACE DISINFECTION AND MEDICAL WASTE

- The contaminated surface environment in hospital rooms is important in the transmission of healthcare-associated pathogens (MRSA, VRE, C. difficile, Acinetobacter).
- Disinfection of noncritical environmental surfaces/equipment is an essential component of Infection prevention.
- Disinfection should render surfaces and equipment free of pathogens in sufficient numbers to cause human disease. Follow CDC D/S guideline.
- When determining the optimal disinfecting product, consider the 5 components (kill claims/time, safety, ease of use, others).
- Comply with federal (OSHA) and state medical waste regulations.
THANK YOU!

www.disinfectionandsterilization.org